Amendments to the Specification:

Please replace the paragraph beginning at line 9, page 59 with the following:

In a preferred embodiment, the nucleic acid molecule encodes a polypeptide that is specific to cancer, a cancer-specific polypeptide (CaSP). In a more preferred embodiment, the nucleic acid molecule encodes a polypeptide that comprises an amino acid sequence of SEQ ID NO: 142-361. In another highly preferred embodiment, the nucleic acid molecule comprises a nucleic acid sequence of SEQ ID NO: 1-141. Nucleotide sequences of the instantly-described nucleic acid molecules were determined by assembling several DNA molecules from either public or proprietary databases. Some of the underlying DNA sequences are the result, directly or indirectly, of at least one enzymatic polymerization reaction (*e.g.*, reverse transcription and/or polymerase chain reaction) using an automated sequencer (such as the ~~MegaBACE™~~ MEGABACE® 1000, Amersham Biosciences, Sunnyvale, CA, USA).

Please replace the paragraph beginning at line 19, page 64 with the following:

Other post-synthetic approaches also permit internal labeling of nucleic acids. For example, fluorophores can be attached using a cisplatin reagent that reacts with the N7 of guanine residues (and, to a lesser extent, adenine bases) in DNA, RNA, and Peptide Nucleic Acids (PNA) to provide a stable coordination complex between the nucleic acid and fluorophore label (Universal Linkage System) (available from Molecular

Probes, Inc., Eugene, OR, USA and Amersham Pharmacia Biotech, Piscataway, NJ, USA); *see* Alers *et al.*, *Genes, Chromosomes & Cancer* 25: 301- 305 (1999); Jelsma *et al.*, *J. NIH Res.* 5: 82 (1994); Van Belkum *et al.*, *BioTechniques* 16: 148-153 (1994). Alternatively, nucleic acids can be labeled using a disulfide-containing linker (~~FastTag™~~ FASTTAG® Reagent, Vector Laboratories, Inc., Burlingame, CA, USA) that is photo- or thermally coupled to the target nucleic acid using aryl azide chemistry; after reduction, a free thiol is available for coupling to a hapten, fluorophore, sugar, affinity ligand, or other marker.

Please replace the paragraph beginning at line 32 of page 86 with the following:

A wide variety of unicellular host cells are useful in expressing the DNA sequences of this invention. These hosts may include well known eukaryotic and prokaryotic hosts, such as strains of, fungi, yeast, insect cells such as *Spodoptera frugiperda* (SF9), animal cells such as CHO, as well as plant cells in tissue culture. Representative examples of appropriate host cells include, but are not limited to, bacterial cells, such as *E. coli*, *Caulobacter crescentus*, *Streptomyces* species, and *Salmonella typhimurium*; yeast cells, such as *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Pichia pastoris*, *Pichia methanolica*; insect cell lines, such as those from *Spodoptera frugiperda* — *e.g.*, Sf9 and Sf21 cell lines, and expresSF™ cells (Protein Sciences Corp., Meriden, CT, USA) — *Drosophila* S2 cells, and *Trichoplusia ni* ~~High-Five®~~ High Five™ Cells (Invitrogen, Carlsbad, CA, USA); and mammalian cells. Typical mammalian cells

include BHK cells, BSC 1 cells, BSC 40 cells, BMT 10 cells, VERO cells, COS1 cells, COS7 cells, Chinese hamster ovary (CHO) cells, 3T3 cells, NIH 3T3 cells, 293 cells, HEPG2 cells, HeLa cells, L cells, MDCK cells, HEK293 cells, WI38 cells, murine ES cell lines (*e.g.*, from strains 129/SV, C57/BL6, DBA-1, 129/SVJ), K562 cells, Jurkat cells, and BW5147 cells. Other mammalian cell lines are well known and readily available from the American Type Culture Collection (ATCC) (Manassas, VA, USA) and the National Institute of General Medical Sciences (NIGMS) Human Genetic Cell Repository at the Coriell Cell Repositories (Camden, NJ, USA). Cells or cell lines derived from breast, colon, lung, ovarian or prostate tissue are particularly preferred because they may provide a more native post-translational processing. Particularly preferred are human breast, colon, lung, ovarian or prostate cells or human breast, colon, lung, ovarian or prostate cancer cells.

Please replace the paragraph beginning at line 31 of page 87 with the following:

Nucleic acid molecules and vectors may be introduced into prokaryotes, such as *E. coli,* in a number of ways. For instance, phage lambda vectors will typically be packaged using a packaging extract (*e.g.*, ~~Gigapack®~~ GIGAPACK® packaging extract, Stratagene, La Jolla, CA, USA), and the packaged virus used to infect *E. coli*.

Please replace the paragraph beginning at line 3 of page 88 with the following:

Plasmid vectors will typically be introduced into chemically competent or electrocompetent bacterial cells. *E. coli* cells can be rendered chemically competent by treatment, *e.g.*, with $CaCl_2$, or a solution of Mg^{2+}, Mn^{2+}, Ca^{2+}, Rb^{+} or K^{+}, dimethyl sulfoxide, dithiothreitol, and hexamine cobalt (III), Hanahan, *J. Mol. Biol.* 166(4):557-80 (1983), and vectors introduced by heat shock. A wide variety of chemically competent strains are also available commercially (*e.g.*, ~~Epicurian Coli®~~ EPICURIAN COLI® ~~XL10-Gold®~~ XL10-GOLD® Ultracompetent Cells (Stratagene, La Jolla, CA, USA); DH5α competent cells (Clontech Laboratories, Palo Alto, CA, USA); and TOP10 Chemically Competent E. coli Kit (Invitrogen, Carlsbad, CA, USA)). Bacterial cells can be rendered electrocompetent to take up exogenous DNA by electroporation by various pre-pulse treatments; vectors are introduced by electroporation followed by subsequent outgrowth in selected media. An extensive series of protocols is provided by BioRad (Richmond, CA, USA).

Please replace the paragraph beginning at line 13 of page 119 with the following:

Other fluorophores include, *inter alia*, ~~Alexa Fluor®~~ ALEXA FLUOR® 350, ~~Alexa Fluor®~~ ALEXA FLUOR® 488, ~~Alexa Fluor®~~ ALEXA FLUOR® 532, ~~Alexa Fluor®~~ ALEXA FLUOR® 546, ~~Alexa Fluor®~~ ALEXA FLUOR® 568, ~~Alexa Fluor®~~ ALEXA FLUOR® 594, ~~Alexa Fluor®~~ ALEXA FLUOR® 647 (monoclonal antibody labeling kits available from Molecular Probes, Inc., Eugene, OR, USA), BODIPY dyes, such as BODIPY 493/503, BODIPY FL, BODIPY R6G, BODIPY 530/550, BODIPY TMR, BODIPY 558/568, BODIPY 558/568, BODIPY 564/570, BODIPY 576/589, BODIPY 581/591, BODIPY TR, BODIPY 630/650, BODIPY

650/665, Cascade Blue, Cascade Yellow, Dansyl, lissamine rhodamine B, Marina Blue, Oregon Green 488, Oregon Green 514, Pacific Blue, rhodamine 6G, rhodamine green, rhodamine red, tetramethylrhodamine, Texas Red (available from Molecular Probes, Inc., Eugene, OR, USA), and Cy2, Cy3, Cy3.5, Cy5, Cy5.5, Cy7, all of which are also useful for fluorescently labeling the antibodies of the present invention. For secondary detection using labeled avidin, streptavidin, captavidin or neutravidin, the antibodies of the present invention can usefully be labeled with biotin.

Please replace the paragraph beginning at line 4 of page 132 with the following:

In addition to detection in bodily fluids, the proteins and nucleic acids of the invention are suitable to detection by cell capture technology. Whole cells may be captured by a variety methods for example magnetic separation, U.S. Patent. Nos. 5,200,084; 5,186,827; 5,108,933; 4,925,788, the disclosures of which are incorporated herein by reference in their entireties. Epithelial cells may be captured using such products as ~~Dynabeads®~~ DYNABEADS® or CELLection™ (Dynal Biotech, Oslo, Norway). Alternatively, fractions of blood may be captured, e.g., the buffy coat fraction (50mm cells isolated from 5ml of blood) containing epithelial cells. In addition, cancer cells may be captured using the techniques described in WO 00/47998, the disclosure of which is incorporated herein by reference in its entirety. Once the cells are captured or concentrated, the proteins or nucleic acids are detected by the means described in

the subject application. Alternatively, nucleic acids may be captured directly from blood samples, see U.S. Patent Nos. 6,156,504, 5,501,963; or WO 01/42504, the disclosures of which are incorporated herein by reference in their entireties.

Please replace the paragraph beginning at line 19 of page 142 with the following:

Due to the association of angiogenesis with cancer vascularization there is great need of new markers and methods for diagnosing angiogenesis activity to identify developing tumors and angiogenesis related diseases. Furthermore, great need is also present for new molecular targets useful in the treatment of angiogenesis and angiogenesis related diseases such as cancer. In addition known modulators of angiogenesis such as endostatin or vascular endothelial growth factor (VEGF). Use of the methods and compositions disclosed herein in combination with anti-angiogenesis drugs, drugs that block the matrix breakdown (such as BMS-275291, Dalteparin (~~Fragmin®~~ FRAGMIN®), Suramin), drugs that inhibit endothelial cells (2-methoxyestradiol (2-ME), CC-5013 (Thalidomide Analog), Combretastatin A4 Phosphate, LY317615 (Protein Kinase C Beta Inhibitor), Soy Isoflavone (Genistein; Soy Protein Isolate), Thalidomide), drugs that block activators of angiogenesis (AE-941 (~~Neovastat™~~ NEOVASTAT®; GW786034), Anti-VEGF Antibody (Bevacizumab; ~~Avastin™~~ AVASTIN®), Interferon-alpha, PTK787/ZK 222584, VEGF-Trap, ZD6474), Drugs that inhibit endothelial-specific integrin/survival signaling (EMD 121974, Anti-Anb3 Integrin Antibody (Medi-522; ~~Vitaxin™~~ VITAXIN®)).

Please replace the paragraph beginning at line 9 of page 401 with the following:

Chromosomes are counterstained with 4,6-diamino-2-phenylidole and propidium iodide, producing a combination of C-and R-bands. Aligned images for precise mapping are obtained using a triple-band filter set (Chroma Technology, Brattleboro, VT) in combination with a cooled charge-coupled device camera (Photometrics, Tucson, AZ) and variable excitation wavelength filters. Johnson (1991). Image collection, analysis and chromosomal fractional length measurements are performed using the ~~ISee~~ ISEE® Graphical Program System. (Inovision Corporation, Durham, NC.) Chromosome alterations of the genomic region hybridized by the probe are identified as insertions, deletions, and translocations. These alterations are used as a diagnostic marker for an associated disease.